

Generalized Autoimmunity of ANCA and ASCA Related to Severity of Disease in Autistic Children with GI Disease

A.J. Russo¹, A Krigsman², B Jepson² and Andrew Wakefield²

¹Research Director, Health Research Institute/Pfeiffer Treatment Center, 4575 Weaver Parkway, Warrenville, Illinois 60555. ²Thoughtful House Center for Children, 3001 Bee Caves Road, Austin, Texas, 78746. Email: ajrusso@hriptc.org

Abstract

Aim: To assess serum Anti-Neutrophil cytoplasmic Antibody (ANCA—PR3 and MPO) and Anti-Saccharomyces Cerevisiae Antibody (ASCA) levels in autistic children with severe gastrointestinal (GI) disease and to test the hypothesis that there is generalized autoimmunity in a subpopulation of autistic children with severe GI disease and that this autoimmunity is associated with the severity of GI disease.

Subjects and Methods: Serum from 40 autistic children with chronic digestive disease (most with ileo-colonic lymphoid nodular hyperplasia (LNH) and inflammation of the colorectum, small bowel and/or stomach), and 24 controls (13 age matched autistic children with no GI disease and 11 age matched children without autism or GI disease) were tested using ELISAs designed to quantitate ANCA (PR3 and MPO) and ASCA levels. ANCA and ASCA concentration of autistic children with GI disease were compared to GI disease severity (including LNH and erythema).

Results: We previously reported that 6 of the 40 autistic children with GI disease in this study had anti-PR3 ANCA. All 6 of these also had anti-MPO IgG. In this study, we found that 6 of the 40 autistic children also had ASCA and 4 of these children with ASCA also had both anti-PR3 and anti-MPO ANCA. These 4 with ANCA and ASCA had significantly higher GI disease severity, particularly associated with LNH and erythema.

Discussion: These results suggest a general autoimmune response (ANCA and ASCA) in a sub group (high GI disease severity) of autistic children with GI disease. The presence of these autoantibodies may be a useful biomarker for autistic children with severe GI disease.

Keywords: autism, autoimmunity, GI disease, ANCA, ASCA

Immunology and Immunogenetics Insights 2009:1 37–47

This article is available from <http://www.la-press.com>.

© the authors, licensee Libertas Academica Ltd.

This is an open access article distributed under the terms of the Creative Commons Attribution License (<http://www.creativecommons.org/licenses/by/2.0>) which permits unrestricted use, distribution and reproduction provided the original work is properly cited.



Introduction

ANCA were originally detected in serum from patients with Wegener's granulomatosis (WG),¹ a disease characterized by necrotizing granulomatous inflammation of the upper and lower airways in conjunction with systemic vasculitis and necrotizing crescentic glomerulonephritis.² Following the detection of ANCA in systemic vasculitis, it became clear that ANCA also occurred in other idiopathic inflammatory disorders,³ including inflammatory bowel diseases or IBD (which include ulcerative colitis (UC) and Crohn's disease (CD)),^{4,5} in autoimmune-mediated liver diseases,^{6–8} in rheumatoid arthritis (RA),^{9,10} and in systemic lupus erythematosus (SLE).^{11,12}

ASCA of IgG and IgA isotype are present in 60% of cases with diagnosed CD,¹³ and directed against phosphopeptidomannans present in the yeast (*S. cerevisiae*) cell walls.^{13,16} ASCA may be elicited due to molecular mimicry and priming by a high mannose-containing bacterial or viral antigen or an "auto-antigenic (self) molecule".¹⁴ It has been observed that levels of ASCA in CD cases are independent of disease activity, duration and/or treatment.¹⁵

Autistic Spectrum Disorder (ASD) is a neurodevelopmental syndrome with onset prior to age 36 months. Diagnostic criteria consists of impairments in sociality and communication plus repetitive and stereotypic behaviors.¹⁷ Traits strongly associated with autism include movement disorders and sensory dysfunctions.¹⁸ Although autism may be apparent soon after birth, many autistic children experience at least several months, up to a year or more in some cases, of normal development—followed by regression, defined as loss of function or failure to progress.^{19–21}

Children with autistic spectrum disorders (ASD) frequently have accompanying gastrointestinal (GI) symptoms and pathology,^{22,23} which includes inflammation of the GI tract.^{24–27} Many autistic children, particularly those with GI disease, also have a higher propensity for fungal infections.^{28,29}

We previously reported that a significant number of autistic children with GI disease have ANCA (both anti-PR3 and anti-MPO), and that there is a relationship between individuals with ANCA and severity of intestinal disease.³⁰ We have also reported that a significant number of autistic children with chronic digestive disease have anti-PR3 ANCA, low

serum AAT, and high serum PR3, which correlate with high severity of GI disease, suggesting that high PR3 levels may be causing ANCA in autistic children with severe GI problems.³¹ Because of this, we hypothesized that the autoimmune response in this sub group of autistic children might be general, so we tested the same population for ASCA.

Materials and Methods

ELISA to measure serum ANCA (anti-PR3 antibodies and anti-MPO antibodies) and ASCA (IMMCO Diagnostics, Buffalo, N.Y.)

All reagents and specimens were equilibrated to room temperature before the assay was performed. A 1:51 dilution of the patient samples were prepared by mixing 10 μ l of the patient's sera with 0.5 ml of Serum Diluent. One hundred microliters of calibrators (20–200 Eu/ml antibodies), positive and Negative control serums, serum diluent alone, and diluted patient samples were added to the appropriate microwells of a microculture plate (each well contained approximately 150 ng of purified PR3 or MPO). Wells were incubated for 30 minutes (\pm 5 min) at room temperature, then washed 4 \times with wash buffer. One hundred microliters of pre-diluter anti-human IgG conjugated with alkaline phosphatase was added to all microwells, incubated for 30 minutes (\pm 5 min) at room temperature, then wash 4 \times with wash buffer. One hundred microliters of enzyme substrate was added to each microwell. After approximately 30 minutes at room temperature, the wells were read at 405 nm with an ELISA reader (BioRad Laboratories, Inc., Hercules, CA, USA).

Subjects and scoring of severity of GI disease

Serum from autistic individuals with GI disease was obtained from the Thoughtful House, Austin, Texas. All 40 children in this study with ASD (median age 6 years; range 2–16; 34 male) with gastrointestinal symptoms, were investigated by ileo-colonoscopy. Macroscopic and histological features of the upper and lower GI tract were scored. A point system was developed to assess the *severity of GI disease* (particularly inflammation). Patients were scored according to mild (1 point), moderate (2 points) and marked (3 points)



disease in each area (upper and lower GI) and for scope (macroscopic) and histology of each area. Therefore, the maximum score for GI disease was 12 (3 points each for upper scope, upper histology, lower scope and lower histology). A point system was also developed for severity of lymphoid nodular hyperplasia (LNH). Patients were scored according to mild (1 point), moderate (2 points) and marked (3 points) LNH in each area (upper and lower GI) for a maximum of 6 points. And finally, a point system was also developed for severity of erythema. Patients were scored according to mild (1 point), moderate (2 points) and marked (3 points) erythema in each area (upper and lower GI) for a maximum of 6 points.

Controls

Two control groups (Total $n = 24$) were studied, including 12 age matched autistic children with no GI disease and 12 age matched children without autism or GI disease (mean 68 months), gender (76% male) and diagnosis (55% regressive onset) matched autistic children with no GI disease; 11 age (mean 72 months) and gender (80% male) matched children without autism or GI disease. Serum and medical history of controls were obtained from the Autism Genetic Resource Exchange—AGRE*.

Serums

Experimental (Thoughtful House) and control (AGRE) serums were frozen at -70 C immediately after collection and cell/serum separation, then stored at -70 C until thawed for use in ELISAs.

Statistics

Inferential statistics were derived from t-test and odds ratios with 95% confidence intervals. ANOVA analysis was used to determine correlation between types of autoantibodies present and severity of GI disease.

Results

Using the ELISAs described above, autistic children with chronic digestive disease, and controls, were

tested for serum ANCA (PR3 and MPO) and ASCA levels. Results of a typical ASCA assay are shown in Figure 1. In each assay, positive titers of autoantibodies were determined by comparing experimental and control serum levels with ASCA standards and negative controls (sample diluent alone) (Fig. 2). The Optical Density of the 20 EU standard was the cutoff point necessary to establish a positive titer. For each assay, there were 3 or 4 replicate samples tested in each group (control and experimental), and each assay was repeated at least twice.

We found that 6 of the 40 autistic children with GI disease had positive ASCA compared to only one of the controls (autistic and non autistic, age and diagnosis matched children without GI disease) (Table 1) ($p < 0.05$).

Four of the 6 with ASCA also had both anti-PR3 and anti-MPO ANCA, compared to none of the controls (Table 2) ($p < 0.01$).

The 4 children with both ANCA and ASCA had significantly higher total GI disease severity ($p \leq 0.005$), including severity of erythema ($p \leq 0.001$) compared to those without the 3 types of autoantibodies. Although not significant, there was also a difference in severity of LNH between the two groups ($p \leq 0.1$) (Tables 3 and 4).

We also found that when ANCA (anti-MPO and anti-PR3) as well as ASCA are present (compared to anti-MPO alone, anti-MPO and antiPR3 alone) there is a higher correlation to high severity GI disease in autistic children when considering Total GI disease ($p < 0.08$) and the presence and severity of LNH ($p < 0.02$) (Table 5).

Discussion

Immune dysfunction has been reported in autistic children including autoimmunity to central nervous system (CNS) proteins.^{32–34} This has led to speculation that exposure of the developing neuronal system during critical periods of aberrant immune activation may result in the brain pathology of ASD. Neuroactive compounds, that share immunomodulatory properties, have been implicated in the disease process. For example, elevated platelet serotonin levels are observed in approximately one third of children with autism.^{43–45} Analysis of data from small but representative groups of ASD patients has shown that approximately 30%–70% of autistic patients have circulating anti-brain

*The Autism Genetic Resource Exchange (AGRE) is the first collaborative gene bank for the study of autism spectrum disorders and one of the world's largest shared resources for the study of autism and related disorders, with a collection of over 900 well-characterized multiplex and simplex families made available to the greater scientific community. Founded by Cure Autism Now (CAN) in 1997, AGRE is currently funded by the National Institute of Mental Health (NIMH) and Autism Speaks (AS), which merged with CAN in 2006.

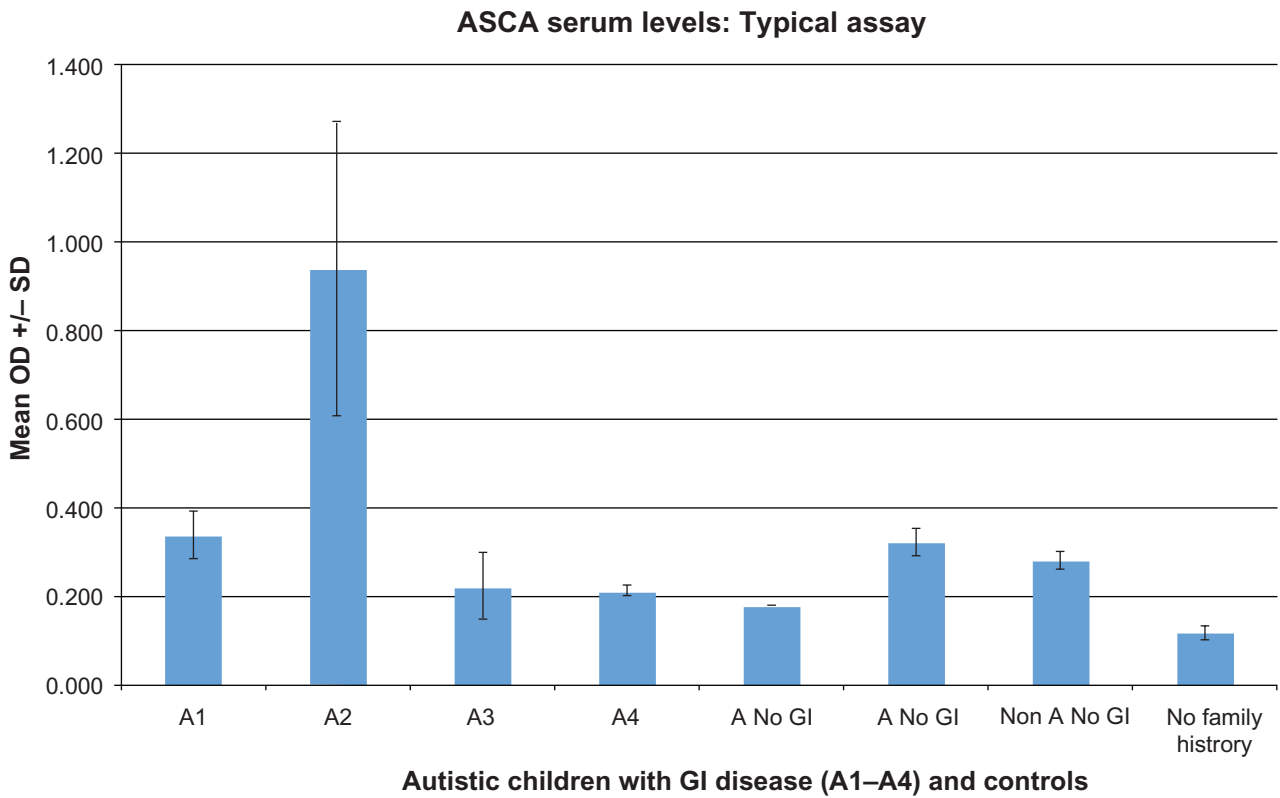


Figure 1. Serum ASCA concentration was measured in a typical ELISA. Four autistic children (A) with GI disease, 2 autistic children with no GI disease controls (A No GI), 1 non-autistic child with no GI disease controls (Non A No GI) and 1 child with no family history of autism with no GI disease (No Family History) were tested. Four replicate samples were tested for each individual.

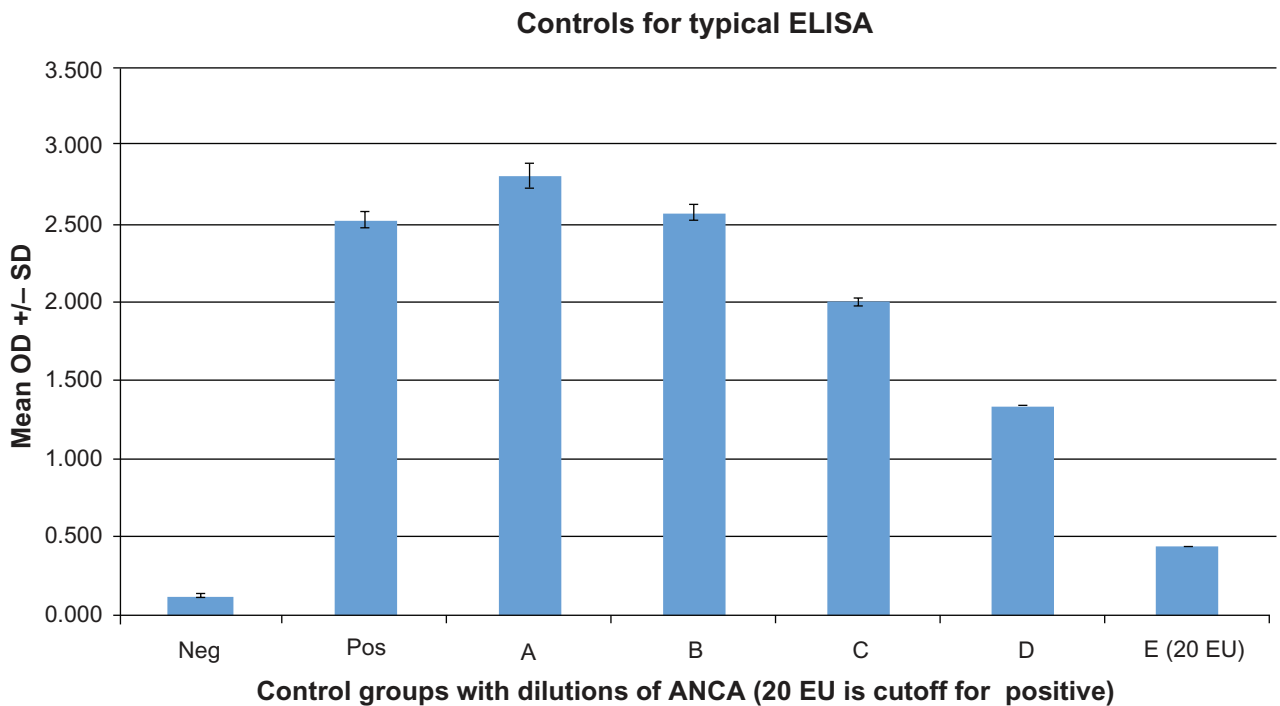


Figure 2. ASCA serum concentration was established for each individual by testing and correlating to known standards of various ASCA titers (A-E) as well as a positive (Serum with known high ASCA) and a negative control (serum diluent alone). Any serums with OD above the OD of the 20 EU (E) control were considered positive.



Table 1. ASCA data.

Autistic children with GI disease (1)					Controls no GI (2)				
Diagnosis	Assay 1	Assay 2	Mean*	SD		Assay 1	Assay 2	Mean*	SD
1 RA	0.16	0.247	0.204	0.062	1 A	0.176		0.176	
2 A	0.147	0.184	0.166	0.026	2 A	0.306		0.306	
3 A	0.286	0.274	0.280	0.008	3 A	0.375		0.375	
4 R-PDD	0.219	0.286	0.253	0.047	4 A	0.187		0.187	
5 RA	0.351	0.447	0.399	0.068	5 A	0.21		0.210	
6 RA	0.556		0.556		6 A-PDD	0.417	0.278	0.348	0.098
7 RA	0.904	0.537	0.721	0.260	7 A-PDD	0.203	0.194	0.199	0.006
8 RA	0.157	0.172	0.165	0.011	8 AR	0.178	0.18	0.179	0.001
9 R-UD	0.31	0.24	0.275	0.049	9 AR	0.343	0.3	0.322	0.030
10 RA	0.176	0.21	0.193	0.024	10 AR	0.223	0.178	0.201	0.032
11 A	0.162	0.188	0.175	0.018	11 AR	0.374	0.35	0.362	0.017
12 RA	0.161	0.185	0.173	0.017	12 AR	0.178	0.229	0.204	0.036
13 PDD/NOS	0.171	0.164	0.168	0.005	13 NA	0.25		0.250	
14 A	0.169	0.195	0.182	0.018	14 NA	0.907		0.907	
15 A	0.464	0.49	0.477	0.018	15 NA	0.382		0.382	
16 RA	0.299	0.374	0.337	0.053	16 NA	0.193	0.227	0.210	0.024
17 A	0.704	1.174	0.939	0.332	17 NA	0.264	0.264	0.264	0.000
18 RA	0.169	0.276	0.223	0.076	18 NA	0.281		0.281	
19 R-PDD	0.426	0.376	0.401	0.035	19 NA	0.197	0.228	0.213	0.022
20 A	0.793		0.793		20 NA	0.159	0.183	0.171	0.017
21 R-PDD	0.228	0.318	0.273	0.064	21 NA	0.197	0.267	0.232	0.049
22 RA	0.196	0.277	0.237	0.057	22 NA	0.23	0.176	0.203	0.038
23 RA	0.277	0.348	0.313	0.050	23 NA	0.24	0.306	0.273	0.047
24 A	0.352	0.318	0.335	0.024	24 NA	0.197	0.224	0.2105	0.019
25 RA	0.151	0.191	0.171	0.028					
26 RA	0.248	0.325	0.287	0.054					
27 R-ASP	0.128	0.153	0.141	0.018					
28 A	0.174	0.235	0.205	0.043					
29 PDD	0.253	0.207	0.230	0.033					
30 R-PDD/NOS	0.343	0.305	0.324	0.027					
31 RA	0.29	0.29	0.290	0.000					
32 A	0.274	0.39	0.332	0.082					
33 RA	0.78	0.611	0.696	0.120					
34 A	0.19	0.205	0.198	0.011					
35 RA	0.169	0.427	0.298	0.182					
36 RA	0.249	0.415	0.332	0.117					
37 RA	0.169	0.208	0.189	0.028					
38 A	0.195	0.218	0.207	0.016					
39 RA	0.169	0.199	0.184	0.021					
40 RA	0.282	0.375	0.329	0.066					

*>0.45 O.D. = positive ASCA >= 20 Eu/ml.

(1) Serum from Thoughtful House, Austin, Texas.

(2) Serum from Autistic Family Members from the Autism Genetic Resource Exchange.

Mean OD and separate data from 2 assays measuring ASCA of 40 autistic children with chronic digestive disease (most with ileo-colonic lymphoid nodular hyperplasia (LNH) and inflammation of the colorectum, small bowel and/or stomach) (Autistic GI), and 24 controls (12 age matched autistic children with no GI disease (A) and 12 age matched non autistic children, without GI disease (NA)).

**Table 2.** ASCA ANCA comparison.

	Mean O.D. anti-PR3**	Mean O.D. anti-MPO***	Mean O.D. ASCA****		Mean O.D. anti-PR3**	Mean O.D. anti-MPO***	Mean O.D. ASCA****
Autistic individuals with GI disease (1)				Controls (Autistic and nonautistic-no GI disease) (2)			
RA	0.231	0.546	0.16	A	0.316	0.377	0.375
A	0.140	0.341	0.147	A	0.351	0.298	0.21
A	0.214	0.620	0.286	A	0.296	0.424	0.264
R-PDD	0.245	0.416	0.219	A	0.264	0.267	0.23
RA	0.253	0.553	0.351	A	0.303	0.585	0.907
RA	0.366	0.450	0.556	A-PDD	0.403	0.453	0.343
RA	0.466	0.833	0.904	A-PDD	0.353	0.215	0.159
RA	0.134	0.274	0.157	AR	0.236	0.224	0.187
R-UD	0.251	0.351	0.31	AR	0.338	0.718	0.178
RA	0.147	0.302	0.176	AR	0.293	0.404	0.193
A	0.129	0.322	0.162	AR	0.343	0.393	0.281
RA	0.121	0.286	0.161	AR	0.311	0.374	0.197
PDD/NOS	0.181	0.317	0.171	AR	0.392	0.298	0.178
A	0.189	0.341	0.169	NA	0.339	0.306	0.176
A	0.401	0.630	0.464	NA	0.314	0.374	0.306
RA	0.196	0.360	0.299	NA	0.284	0.272	0.417
A	0.179	0.379	0.704	NA	0.325	0.365	0.203
RA	0.175	0.329	0.169	NA	0.336	0.337	0.223
R-PDD	0.443	0.539	0.426	NA	0.298	0.394	0.374
A	0.576	0.629	0.793	NA	0.4	0.38	0.25
R-PDD	0.278	0.529	0.228	NA	0.424	0.457	0.382
RA	0.199	0.503	0.196	NA	0.312	0.356	0.197
RA	0.544	0.504	0.277	NA	0.241	0.229	0.24
A	0.218	0.387	0.352	NA	0.247	0.381	0.197
RA	0.140	0.356	0.151				
RA	0.259	0.463	0.248				
R-ASP	0.099	0.225	0.128				
A	0.174	0.457	0.174				
PDD	0.197	0.361	0.253				
R-PDD/NOS	0.274	0.506	0.343				
RA	0.235	0.435	0.29				
A-dev. plateau	0.233	0.371	0.274				
RA	0.482	0.566	0.78				
A	0.136	0.284	0.19				
RA	0.227	0.306	0.169				
RA	0.221	0.537	0.249				
RA	0.141	0.414	0.169				
A	0.173	0.394	0.195				
RA	0.166	0.313	0.169				
RA	0.289	0.327	0.282				

****>0.45 O.D. = positive ASCA > = 20 Eu/ml.

***>0.5 O.D. = positive anti-MPO antibodies > = 20 Eu/ml.

** >0.4 O.D. = positive Anti-PR3 antibodies > = 20 Eu/ml.

(1) Serum from Thoughtful House, Austin, Texas.

(2) Serum from Autistic Family Members from the Autism Genetic Resource Exchange.

Mean OD and separate data from 2 assays showing comparison of ASCA and ANCA data of 40 autistic children with chronic digestive disease (most with ileo-colonic lymphoid nodular hyperplasia (LNH) and inflammation of the colorectum, small bowel and/or stomach) (Autistic GI), and 24 controls (12 age matched autistic children with no GI disease (A) and 12 age matched non autistic children, without GI disease (NA)).

**Table 3.** Autoimmunity compared to severity of disease.

	Mean O.D. anti-PR3**	Mean O.D. anti-MPO***	Mean O.D. ASCA****	Diagnosis	LNH	Eryth	Total GI
Autistic individuals with GI disease							
1	0.231	0.546	0.16	RA	2	0	5
2	0.140	0.341	0.147	A	3	0	6
3	0.214	0.620	0.286	A	3	6	9
4	0.245	0.416	0.219	R-PDD	1	0	3
5	0.253	0.553	0.351	RA	1	0	3
6	0.366	0.450	0.556	RA	2	2	6
7	0.466	0.833	0.904	RA	4	3	8
8	0.134	0.274	0.157	RA	2	0	8
9	0.251	0.351	0.31	R-UD	4	2	8
10	0.147	0.302	0.176	RA	3	1	7
11	0.129	0.322	0.162	A	3	1	6
12	0.121	0.286	0.161	RA	1	0	7
13	0.181	0.317	0.171	PDD/NOS	2	2	6
14	0.189	0.341	0.169	A	1	2	6
15	0.401	0.630	0.464	A	3	5	10
16	0.196	0.360	0.299	RA	0	2	5
17	0.179	0.379	0.704	A	2	1	5
18	0.175	0.329	0.169	RA	3	1	NA
19	0.443	0.539	0.426	R-PDD	5	0	NA
20	0.576	0.629	0.793	A	4	4	11
21	0.278	0.529	0.228	R-PDD	3	1	8
22	0.199	0.503	0.196	RA	3	2	5
23	0.544	0.504	0.277	RA	3	0	7
24	0.218	0.387	0.352	A	3	2	8
25	0.140	0.356	0.151	RA	2	0	4
26	0.259	0.463	0.248	RA	3	0	4
27	0.099	0.225	0.128	R-ASP	2	1	6
28	0.174	0.457	0.174	A	3	4	6
29	0.197	0.361	0.253	PDD	2	1	4
30	0.274	0.506	0.343	R-PDD/NOS	2	1	4
31	0.235	0.435	0.29	RA	3	0	6
32	0.233	0.371	0.274	A-dev. plateau	2	0	4
33	0.482	0.566	0.78	RA	3	2	6
34	0.136	0.284	0.19	A	0	0	NA
35	0.227	0.306	0.169	RA	3	0	5
36	0.221	0.537	0.249	RA	2	2	7

(Continued)



Table 3. (Continued)

	Mean O.D. anti-PR3**	Mean O.D. anti-MPO***	Mean O.D. ASCA****	Diagnosis	LNH	Eryth	Total GI
Autistic individuals with GI disease							
37	0.141	0.414	0.169	RA	2	0	3
38	0.173	0.394	0.195	A	4	0	7
39	0.166	0.313	0.169	RA	3	2	7
40	0.289	0.327	0.282	RA	6	0	10

***>0.5 O.D. = positive anti-MPO antibodies >= 20 Eu/ml.

** >0.4 O.D. = positive anti-PR3 antibodies >= 20 Eu/ml.

****>0.45 O.D. = positive ASCA >= 20 Eu/ml.

Scale: 1-mild, 2-moderate, 3-severe, I-intestine, C-colon, N-normal, NA information not available, HC-hypovascularization.

Diagnosis: A-autistic, RA-regression, ASP-Asperger's, PDD-pervasive developmental disorder.

Total GI score = sum of US, UH, LS, LH scores.

Mean OD and separate data from 2 assays showing comparison of ASCA and ANCA data of 40 autistic children with chronic digestive disease (most with ileocolonic lymphoid nodular hyperplasia (LNH) and inflammation of the colorectum, small bowel and/or stomach) compared to the severity of disease (below). Diagnosis of autistic children with GI disease (A-autistic; RA-autistic with regressive onset; PDD-pervasive developmental disorder; UD-undetermined; ASP-aspergers) and severity of lymphonodular hyperplasia (LNH), presence and severity of erythema and total GI disease severity is posted.

Patients were scored according to mild (1 point), moderate (2 points) and marked (3 points) disease in each area (upper and lower GI) and for scope (macroscopic) and histology of each area. Therefore the maximum score for GI disease would be 12 (3 points each for upper scope, upper histology, lower scope and lower histology). A point system was also developed for severity of lymphoid nodular hyperplasia (LNH). Patients were scored according to mild (1 point), moderate (2 points) and marked (3 points) LNH in each area (upper and lower GI) for a maximum of 6 points. And finally, a point system was also developed for severity of erythema. Patients were scored according to mild (1 point), moderate (2 points) and marked (3 points) erythema in each area (upper and lower GI) for a maximum of 6 points.

Abbreviations: US, upper scope; UH, upper histology; LS, lower scope; LH, lower histology; LNH, lymphoid nodular hyperplasia.

Table 4. Autoimmunity vs. severity of disease.

Group	Total GI no auto	Total GI autoantibodies
Mean	5.91	8.75
SD	1.76	2.22
SEM	0.31	1.11
N	33	4
t test		
p <= 0.0052		
	LNH no auto	LNH autoantibodies
Mean	2.47	3.5
SD	1.21	0.58
SEM	0.2	0.29
N	36	4
t test		
p <= 0.1		
	Erythema no auto	Erythema autoantibodies
Mean	1	3.5
SD	1.31	1.29
SEM	0.22	0.65
N	36	4
t test		
p <= 0.001		

Children with all three autoantibodies (anti-PR3, anti-MPO and ASCA) were compared to children without these three autoantibodies with respect to Severity of disease (Total GI), severity of LNH, and severity of erythema. T test values of each comparison is shown.

autoantibodies, including autoantibodies to a serotonin receptor,³⁶ myelin basic protein³⁵ and unknown antigens from adult brain tissue extract.⁴⁶

Separate epidemiologic studies suggest that a family history of autoimmune disorders is more common among children with autism than healthy control children.^{38,39} There is also increased incidence of asthma, allergy, autoimmune psoriasis and Type I diabetes in mothers of children with ASD,⁴⁰ and first degree relatives of children with autism and Aspergers are more likely to have an autoimmune disease compared to controls.^{41,42}

While significantly higher levels of autoantibodies are detected in autistic patients when compared with controls, the pathophysiological significance of these antibodies is uncertain. Moreover, their presence has not correlated with an anticipated pathological effect. For example, autoantibodies to myelin basic protein are frequently detected in autistic children, but no signs of associated demyelination have been described,⁴⁷ and anti-brain autoantibodies have also been found in patients with neurological disorders other than autism, as well as in normal individuals. This has raised the question of pathogenic significance and disease-specificity of the antibodies found in the serum of patients with autism, hypothesizing that the



Table 5. Relationship between autoantibody combination and severity of disease.

Autistic GI	Total GI				LNH				Erythema			
	MPO	MPO/PR3	AII	ASCA	MPO	MPO/PR3	AII	ASCA	MPO	MPO/PR3	AII	ASCA
1 RA	5				2				0			
2 A												
3 A	9				3				6			
4 R-PDD												
5 RA	3				1				0			
6 RA				6				2				2
7 RA			8				4				3	
8 RA												
9 R-UD												
10 RA												
11 A												
12 RA												
13 PDD/NOS												
14 A												
15 A			10				3				5	
16 RA												
17 A	5			5	2			2	2			1
18 RA												
19 R-PDD												
20 A			11				4				4	
21 R-PDD	8				3				1			
22 RA	5				3				2			
23 RA		7				3				0		
24 A												
25 RA												
26 RA												
27 R-ASP												
28 A												
29 PDD												
30 R-PDD/NOS	4				2				1			
31 RA												
32 A-dev. plateau												
33 RA			6				3				2	
34 A												
35 RA												
36 RA	7				2				2			
37 RA												
38 A												
39 RA												
40 RA												
Mean	5.75	7.00	8.75	5.50	2.25	3.00	3.50	2.00	1.75	0.00	3.50	1.50
SD	1.92	0.00	1.92	0.71	0.66	0.00	0.50	0.00	1.79	0.00	1.12	0.50
SEM	0.73	0.00	1.11	0.50	0.19	0.00	0.47	0.00	0.17	0.00	0.47	0.61
P			0.08				0.02				0.18	

We compared the severity of disease of autistic children with GI disease with the presence of autoantibodies. When ANCA (anti-MPO and anti-PR3) as well as ASCA are present (compared to anti-MPO alone, anti-MPO and anti-PR3 alone) there is a higher correlation to high severity GI disease in autistic children when considering Total GI disease ($p < 0.08$) and the presence and severity of LNH ($p < 0.02$).



production of these antibodies may be secondary to innate CNS pathology and may simply be a marker of an event in the CNS that allows for the presentation of self-antigens.⁴⁸

An association with autoimmune enteropathies with specific antibodies targeted to gut epithelial cells has been shown in ASD,⁴⁹ and two separate epidemiologic studies suggest that a family history of autoimmune disorders is more common among children with autism than healthy control children.^{50,51}

Up to this point, however, it has been unclear whether individual autistic children are positive for more than one antibody. We have previously shown that a subset of autistic children with GI disease have ANCA.^{30,31}

The data presented here demonstrates that a small sub group of autistic children with severe GI disease have ASCA and most of these individuals also have ANCA to both PR3 and MPO. These results also suggest that this group has a generalized autoimmunity, which includes antibodies to PR3, MPO and *S. cerevisiae*, and, in combination, these antibodies are related to the severity of their GI disease, suggesting that autistic children with these three autoantibodies may be susceptible to the most severe GI disease that the presence of these antibodies may be a marker for this sub group of autistic children.

Disclosures

The authors report no conflicts of interest.

References

1. Van der Woude FJ, Rasmussen N, Lobatto S, et al. Autoantibodies against neutrophils and monocytes: tool for diagnosis and marker of disease activity in Wegener's granulomatosis. *Lancet*. 1985;1(8426):425-9.
2. Kallenberg CGM, Brouwer E, Weening JJ, Cohen Tervaert JW. Antineutrophil cytoplasmic antibodies: current diagnostic and pathophysiological potential. *Kidney Int*. 1994;46:1-15.
3. Kallenberg CGM, Mulder AHL, Cohen Tervaert JW. Antineutrophil cytoplasmic antibodies: a still-growing class of autoantibodies in inflammatory disorders. *Am J Med*. 1992;93:675-82.
4. Saxon A, Shanahan F, Landers CJ, et al. A distinct subset of antineutrophil cytoplasmic antibodies is associated with inflammatory bowel disease. *J Allergy Clin Immunol*. 1990;86:202-10.
5. Rump JA, Schölmerich J, Gross V, et al. A new type of perinuclear antineutrophil cytoplasmic antibody (p-ANCA) in active ulcerative colitis but not in Crohn's disease. *Immunobiol*. 1990;181:406-13.
6. Duerr RH, Targan SR, Landers CJ, et al. Neutrophil cytoplasmic antibodies: a link between primary sclerosing cholangitis and ulcerative colitis. *Gastroenterol*. 1991;100:1385-91.
7. Mulder AHL, Horst G, Haagsma EB, et al. Prevalence and characterization of neutrophil cytoplasmic antibodies in autoimmune liver diseases. *Hepatology*. 1993;17:411-7.
8. Roozendaal C, Van Milligen de Wit AWM, Haagsma EB, et al. Antineutrophil cytoplasmic antibodies in primary sclerosing cholangitis: defined specificities may be associated with distinct clinical features. *Am J Med*. 1998;105:393-9.
9. Lassoued S, Sixou L, Oksman F, et al. Antineutrophil cytoplasmic antibodies and antibodies to myeloperoxidase in rheumatoid arthritis. *Arthritis Rheum*. 1991;34:1069-70.
10. Mulder AHL, Horst G, Van Leeuwen MA, et al. Anti-neutrophil cytoplasmic antibodies in rheumatoid arthritis: characterization and clinical correlates. *Arthritis Rheum*. 1993;36:1054-60.
11. Schnabel A, Csernok E, Isenberg DA, et al. Antineutrophil cytoplasmic antibodies in systemic lupus erythematosus: prevalence, specificities, and clinical significance. *Arthritis Rheum*. 1995;38:633-7.
12. Spronk PE, Bootsma H, Horst G, et al. Antineutrophil cytoplasmic antibodies in systemic lupus erythematosus. *Br J Rheumatol*. 1996;35:625-31.
13. Sutton CL, Yang H, Li Z, et al. Familial expression of anti-Saccharomyces cerevisiae mannan antibodies in affected and unaffected relatives of patients with Crohn's disease. *Gut*. 2000;46:58-63.
14. McKay DM. Bacterial superantigens: Provocateurs of gut dysfunction and inflammation. *Trends Immunol*. 2001;22:497-501.
15. Hoffenberg EJ, Fidanza S, Sauaia A. Serologic testing for inflammatory bowel disease. *J Pediatr*. 1999;134:447-52.
16. Peeters M, Joossens S, Vermeire S, et al. Diagnostic value of anti-Saccharomyces cerevisiae and antineutrophil cytoplasmic autoantibodies in inflammatory bowel disease. *Am J Gastroenterol*. 2001;96:730-4.
17. American Psychiatric Association. Diagnostic and Statistical Manual of Mental Disorders, 4th edition. Washington D.C.: American Psychiatric Association. 1994.
18. Gillberg C, Coleman M. The Biology of the Autistic Syndromes, 2nd edn. London: Mac Keith Press. 1992.
19. Filipek P, Accardo P, Baranek G, et al. The screening and diagnosis of autistic spectrum disorders. *J Autism Dev Disord*. 1999;29(6):439-84.
20. Bailey A, Phillips W, Rutter M. Autism: towards an integration of clinical, genetic, neuro-psychological, and neurobiological perspectives. *J Child Psychol Psychiatry*. 1996;37(1):89-126.
21. Horvath K, Perman J. Autistic disorder and gastrointestinal disease. *Curr Opin Pediatr*. 2002;14(5):583-7.
22. Molloy C, Manning-Courtney P. Prevalence of chronic gastrointestinal symptoms in children with autism and autistic spectrum disorders. *Autism*. 2003;7(2):165-71.
23. Valicenti-McDermott M, McVicar K, Rapin I. Frequency of gastrointestinal symptoms in children with autistic spectrum disorders and association with family history of autoimmune disease. *J Dev Behav Pediatr*. 2006; 27(Suppl 2):S128-36.
24. Ashwood P, et al. Intestinal lymphocyte populations in children with regressive autism: evidence for extensive mucosal immunopathology. *J Clin Immunol*. 2003 Nov; 23(6):504-17.
25. Balzola F, et al. Autistic enterocolitis: confirmation of a new inflammatory bowel disease in an Italian cohort of patients. *Gastroenterology*. 2005;128 Suppl 2:A-303.
26. Wakefield AJ, et al. Review article: the concept of entero-colonic encephalopathy, autism and opioid receptor ligands. *Aliment Pharmacol Ther*. 2002;16(4):663-74.
27. Wakefield AJ, et al. Enterocolitis in children with developmental disorders. *Am J Gastroenterol*. 2000;95(9):2285-95.
28. Shaw W, Kassen E, Chaves E. Increased urinary excretion of analogs of Krebs cycle metabolites and arabinose in two brothers with autistic features. *Clin Chem*. 1995;41:1094-104.
29. Shaw W, Kassen E, Chaves E. Assessment of antifungal drug therapy in autism by measurement of suspected microbial metabolites in urine with gas chromatography-mass spectrometry. *Clin Pract Alternat Med*. 2000;1:15-26.
30. Russo AJ, Krigsman A, Jepson B, Wakefield A. Anti-PR3 and Anti-MPO IgG ANCA in Autistic Children With Chronic GI Disease. *Immunology and Immunogenetics Insights*. 2009;2:21-8.
31. Russo AJ, Krigsman A, Jepson B, Wakefield A. Low Serum Alpha-1 Antitrypsin Associated With Anti-PR-3 ANCA in Autistic Children With GI Disease, accepted for publication. *Genomics Insights*. 2009;2:1-9.



32. Trottier G, Srivastava L, Walker CD. Etiology of infantile autism: a review of recent advances in genetic and neurobiological research. *J Psychiatry Neurosci*. 1999;24(2):103–15.
33. Van Gent T, Heijnen CJ, Treffers PD. Autism and the immune system. *J Child Psychol Psychiatry Allied Discip*. 1997;38(3):337–49.
34. Singh VK, Warren R, Averett R, Ghaziuddin M. Circulating autoantibodies to neuronal and glial filament proteins in autism. *Pediatr Neurol*. 1997; 17(1):88–90.
35. Singh VK, Warren RP, Odell JD, Warren WL, Cole P. Antibodies to myelin basic protein in children with autistic behavior. *Brain Behav Immun*. 1993; 7(1):97–103.
36. Singh VK, Singh EA, Warren RP. Hyperserotonemia and serotonin receptor antibodies in children with autism but not mental retardation. *Biol Psychiatry*. 1997;41(6):753–5.
37. Silva SC, Correia C, Fesel C, et al. Autoantibody repertoires to brain tissue in autism nuclear families. *J Neuro Immunol*. 2004;152(12):176–8.
38. Comi AM, Zimmerman AW, Frye VH, Law PA, Peeden JN. Familial clustering of autoimmune disorders and evaluation of medical risk factors in autism. *J Child Neurol*. 1999;14(6):388–94.
39. Sweeten TL, Bowyer SL, Posey DJ, Halberstadt GM, McDougale CJ. Increased prevalence of familial autoimmunity in probands with pervasive developmental disorders. *Pediatrics*. 2003;112(5):e420.
40. Yan J. Autistic Children More Often Have Parent With Mental Illness. *Psychiatr News*. 2008;43(12):22.
41. Sweeten TL, Bowyer SL, Posey DJ, Halberstadt GM, McDougale CJ. Increased prevalence of familial autoimmunity in probands with pervasive developmental disorders. *Pediatrics [Serial Online]*. 2003;112:e420.
42. Croen LA, Grether JK, Yoshida CK, Oduli R, Van de Water J. Maternal autoimmune diseases, asthma and allergies and childhood autism. *Arch Pediatr Adolesc Med*. 2004.
43. Cook EH, Leventhal BL. The serotonin system in autism. *Curr Opin Pediatr*. 1996;8(4):348–54.
44. Cook EH. Autism: review of neurochemical investigation. *Synapse*. 1990;6(3):292–308.
45. Betancur C, Corbex M, Spielwog C, et al. Serotonin transporter gene polymorphisms and hyperserotonemia in autistic disorder. *Mol Psychiatry*. 2002;7(1):67–71.
46. Todd RD, Ciaranello RD. Demonstration of inter- and intraspecies differences in serotonin binding sites by antibodies from an autistic child. *Proc Natl Acad Sci U S A*. 1985;82(2):612–6.
47. Rumsey JM, Ernst M. Functional neuroimaging of autistic disorders. *Ment Retard Dev Disabil Res Rev*. 2000;6(3):171–9.
48. Rosengren LE, Ahlsen G, Belfrage M, Gillberg C, Haglid KG, Hamberger AA. Sensitive ELISA for glial fibrillary acidic protein: application in CSF of children. *J Neurosci Methods*. 1992;44(2–3):113–9.
49. Torrente F, Ashwood P, Day R, et al. Small intestinal enteropathy with epithelial IgG and complement deposition in children with regressive autism. *Mol Psychiatry*. 2002;7(4):375–82.
50. Comi AM, Zimmerman AW, Frye VH, Law PA, Peeden JN. Familial clustering of autoimmune disorders and evaluation of medical risk factors in autism. *J Child Neurol*. 1999;14(6):388–94.
51. Sweeten TL, Bowyer SL, Posey DJ, Halberstadt GM, McDougale CJ. Increased prevalence of familial autoimmunity in probands with pervasive developmental disorders. *Pediatrics*. 2003;112(5):e420.

Publish with Libertas Academica and every scientist working in your field can read your article

“I would like to say that this is the most author-friendly editing process I have experienced in over 150 publications. Thank you most sincerely.”

“The communication between your staff and me has been terrific. Whenever progress is made with the manuscript, I receive notice. Quite honestly, I’ve never had such complete communication with a journal.”

“LA is different, and hopefully represents a kind of scientific publication machinery that removes the hurdles from free flow of scientific thought.”

Your paper will be:

- Available to your entire community free of charge
- Fairly and quickly peer reviewed
- Yours! You retain copyright

<http://www.la-press.com>